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PURIFICATION OF HUMAN TROPONIN I

This application is a continuation of application Serial No. 09/903,398, filed July 10, 2001, and claims priority under 35 U.S.C. §119(e) of provisional application Serial No. 60/217,069, filed July 10, 2000.

FIELD OF THE INVENTION

The present invention is directed to methods for purifying human Troponin I.

BACKGROUND OF THE INVENTION

Troponin I (TnI) is a component of a heterotrimeric complex, along with troponin C (TnC) and troponin T (TnT), involved in regulation of vertebrate striated muscle contraction (Zot and Potter, Annu. Rev. Biophys. Biophys. Chem. 1987, 16:535-559; Farah and Reinah, FASEB J. 1995, 9:755-767). Muscle contraction is triggered by the binding of Ca⁺⁺ ions to TnC. TnT binds to tropomyosin anchoring the Tn to the muscle filament. TnI is the inhibitory subunit of the troponin complex, binding to actin-tropomyosin complexes and preventing the interaction of actin and myosin. TnI is present in muscle tissue in multiple isoforms expressed from a multi-gene family (Wu *et al.*, DNA Seq. J. DNA Seq. Mapp. 1993, 4:113-121).

Recent investigations have demonstrated a second important biological function for TnI, the ability of the molecule to inhibit both *in vitro* endothelial cell survival/proliferation and inhibit *in vivo* angiogenesis (the growth and development of blood vessels) (Moses *et al.*, Proc. Natl.

Acad. Sci. USA 1999, 96:2645-50). TnI that inhibited endothelial cell development was isolated initially from cartilagenous tissue, and subsequent studies demonstrated anti-angiogenic activity with recombinant forms of TnI expressed in *E. coli*.

The TnI subunit is a single polypeptide with a molecular weight of 21,338. The molecule contains three cysteine residues, at positions 45, 65, and 134 (Wilkinson and Grand, Biochem J. 1975, 149:493-496). Purification processes previously developed to isolate native TnI from tissue sources or recombinant TnI require maintenance of a reducing environment throughout the purification and storage of TnI, ordinarily by the addition of dithiothreitol (DTT) to protein preparations (Potter, Methods Enzymol., 1982, 85:241-263; Jha *et al.*, Protein Exp. Purif., 1994, 5:604-613; Al-Hillawi *et al.*, Eur. J. Biochem., 1994, 225:1195-1201). The added DTT maintains the TnI cysteine sidechain sulfhydryls in their reduced state, preventing the formation of intra- or intermolecular disulfide bond crosslinks. Disulfide bonds between TnI cysteine sidechains are not believed to be present in the active conformation of the protein involved in regulation of muscle contraction (Kluwe *et al.*, FEBS Lett., 1993, 323:83-88), and reductant was utilized in the isolation of active anti-angiogenic forms of TnI.

The necessity of maintaining a reducing environment in processing and storage during large scale purification of TnI for commercial use poses numerous technical difficulties and increases production cost. Thus, there is a need in the art to purify and refold Troponin I efficiently and affordably.

SUMMARY OF THE INVENTION

The present invention provides a method of preparing Troponin I. This method comprises protecting sulfhydryl groups of reduced Troponin I, particularly recombinant TnI. In a preferred embodiment of the invention, the free sulfhydryl groups are protected by sulfitolization (*e.g.*, via reaction with sodium tetrathionate) of Troponin I expressed in a bacterial expression system. Protection of the sulfhydryl groups during Troponin I preparation obviates the costly need for maintaining non-reducing conditions throughout protein preparation, purification, and storage.

In a related aspect, the present invention encompasses sulfhydryl-protected Troponin I itself, and in a preferred embodiment, the Troponin I is denatured and the sulfhydryl groups

protected by sulfates.

The present invention also provides a method of purifying Troponin I, which method comprises subjecting recombinant Troponin I comprising sulfhydryl protecting groups to chromatography. In a preferred embodiment of the invention, the sulfhydryl groups are protected by sulfitolysis. In one aspect of the invention, the Troponin I is subjected to chromatography under non-reducing conditions. In a preferred embodiment of the invention the Troponin I to be purified is expressed in a bacterial expression system such as *E.coli*. In another preferred embodiment, the chromatographic support is an anion exchange column, optionally followed by hydrophobic interaction chromatography.

These and other aspects of the invention are more fully examined in the accompanying Drawings, Detailed Description, and Example.

DESCRIPTION OF THE DRAWINGS

Figures 1A and 1B. A. Proposed reaction for oxidative sulfitolysis. B. Cleavage of disulfide bond by sodium sulfite to form the S-sulfo derivative.

Figure 2. Preparation and washing of TnI-containing inclusion bodies.

Figure 3. Summary of rTroponin-I preparation.

Figure 4. Q-Sepharose FF chromatography of Troponin I. Buffer A: 6M urea, 25mM Tris-HCl, pH 7.5, 100mM; Buffer B: 6M urea, 25mM Tris-HCl, pH 7.5, 2M NaCl; Gradient: Step, 0% B for the flow-through and 100% B for the strip; and Flow rate: 150ml/min.

Figure 5. 300ml Q-sepharose FF chromatography. Buffer A: 6M urea, 25mM Tris-HCl, pH 7.5, 100mM; Buffer B: 6M urea, 25mM Tris-HCl, pH 7.5, 2M NaCl; Gradient: Step, 4% B for elution and 50% B for strip; and Flow rate: 20ml/min.

Figure 6. SDS-PAGE analysis troponin lot after anion exchange steps no. 1 and no. 2 in 16% tris-glycine gel, under non-reducing conditions. A-H refer to lanes in the SDS-PAGE gel. A. Sulfitolyzed troponin Lot 3L4 standard; B. solubilized inclusion bodies; C. sulfitolyzed inclusion bodies (AEX No. 1 load); D. anion exchange no. 1 flowthrough; E. anion exchange no. 1 salt eluate; F. anion exchange no. 2 load; G. anion exchange no. 2 flowthrough; and, H. anion exchange no. 2 100mM NaCl eluate.

Figure 7. Toyopearl 650M (phenyl) HIC chromatograph. Buffer A: 6M urea, 25mM Tris-HCl, pH 7.5, 1M (NH₄)₂SO₄; Buffer B: 6M urea, 25mM Tris-HCl, pH 7.5; Gradient: Step, 100% B for the flow-through and 0% B for strip; and Flow rate: 10ml/min.

Figures 8. SDS-PAGE analysis troponin lot after hydrophobic interaction chromatography in 16% tris-glycine gel, under non-reducing conditions. A-F refers to lanes in the SDS-PAGE gel. A. Sulfitolyzed troponin Lot 3L4 standard; B. AEX step no. 2, troponin eulate pool; C. HIC load (w/1M ammonium sulfate); D. HIC flowthrough (troponin product); E. HIC low salt eulate (column strip); F. lot 3L5 sulfitolyzed troponin product.

Figure 9. Quantitation of rTnI on Zorbax C3.

Figure 10. Troponin I LysC mapping.

Figure 11. SDS-PAGE analysis of sulfitolyzed troponin reduction with dithiothreitol for 45 mins. at ambient temperature. One mg/ml TnI in 6M urea, 25mM tris, 0.15M NaCl pH 7.5, run on 16% tris-glycine gel. 1. 10., Mark 12 MW Stds; 2. 9., sulfitolyzed TnI; 3. 0.05 mM DTT; 4. 0.10 mM DTT; 5. 0.2 mM DTT; 6. 0.3 mM DTT; 7. 0.5 mM DTT; 8. 1.0 mM DTT.

DETAILED DESCRIPTION

Troponin I from human cartilage has recently been reported to possess antiangiogenic activity. In order to produce protein to exploit the antiangiogenic properties of recombinant troponin I, we overexpressed a human skeletal troponin I cDNA in *E. coli*. Expression levels ranged from 2-10 mg/gram of wet cell paste. The recombinant troponin I was isolated from the lysed cells in inclusion bodies, which were solubilized and modified by sulfitolization of cysteine residues to improve protein processing. The sulfitolyzed protein was purified from the inclusion bodies by sequential anion exchange and hydrophobic interaction chromatography. Cysteine protecting groups could be removed by reduction prior to final protein formulation. Overall yield of troponin from the multi-step purification was greater than 50% at purity levels of greater than 95%. The purified recombinant human troponin I is structurally characterized, *e.g.*, by LC/MS, peptide mapping, capillary electrophoresis, SEC with laser light scattering detection, and SDS-PAGE.

The present invention provides a method to purify and refold recombinant TnI which obviates the need for reducing agents by utilizing sulfhydryl protecting groups on the reduced

protein, in particular by oxidative sulfitolysis (Chan, Biochemistry, 1968, 7(12):4247-4253) (Figure 1). This embodiment involves an initial modification of the cysteines in troponin to yield stable S-sulfonated sidechains, which are maintained on the protein during processing and storage. The sulfate protecting groups can be removed from the TnI cysteines by treatment with a reductant to regenerate the free cysteine sulfhydryls.

The term "purified" as used herein refers to material that has been isolated under conditions that reduce or eliminate unrelated materials, *i.e.*, contaminants. For example, a purified protein is preferably substantially free of other proteins or nucleic acids with which it is associated in a cell. As used herein, the term "substantially free" is used operationally, in the context of analytical testing of the material. Preferably, purified material substantially free of contaminants is at least 50% pure; more preferably, at least 90% pure, and more preferably still at least 99% pure. Purity can be evaluated by chromatography, gel electrophoresis, immunoassay, composition analysis, biological assay, and other methods known in the art.

Recombinant TnI can be expressed in bacterial systems in a soluble form or an insoluble form, in inclusion bodies. Recovery of TnI from inclusion bodies requires treatment with solubilizing protein denaturants like urea. In addition, TnI, whose theoretical pI is 8.8, has limited solubility at pH values above 4 in the absence of chaotropic agents, although TnI is soluble at levels of 10-20 mg/ml at low pH (less than 3). High levels (1-6M) of the protein denaturant urea are therefore ordinarily maintained during protein purification of TnI to ensure high solubility and good protein recovery.

The term "refolding" means changes in the three-dimensional conformation of the protein, which restore the protein's biological activity, including its antiangiogenic properties.

Protein refolding of TnI into a bioactive conformation in a buffer and at a pH suitable for use in human patients requires removal of solubilizing denaturant. Circular dichroism and fluorescence studies of TnI protein folding have shown a non-linear relationship relative to chaotrope concentration in solution, with little or no apparent secondary structure at high urea concentration, and putative intermediate folded state(s) occurring between 2-4M urea (Morjana and Tal, Biotechnol. Appl. Biochem., 1998, 28:7-17). This protein refolding process can be accomplished by dilution of TnI in a denaturant solution to an appropriate concentration, generally

below 10 nanomolar, followed by removal of solubilizing chaotrope by dialysis, diafiltration, or gel filtration. Proper refolding requires appropriately timed transit through the chaotrope concentrations of 2-4M urea, where protein folding intermediates are formed, followed by a final protein concentration step.

As used herein, the term "recombinant TnI" (or troponin I) refers to TnI prepared by a biological fermentation process. TnI is a polypeptide of about 21kD containing three cysteine residues, although the present invention encompasses modified forms of TnI lacking one or two cysteine residues. In a specific embodiment, TnI refers to the protein described by Wilkinson and Grand (Biochem. J. 1975, 149:493-496). In a preferred embodiment, TnI of the invention lacks any disulfide bonds.

Recombinant Expression

As noted above, TnI can be prepared, albeit with certain difficulties, by fermentation of genetically modified cells. Preferably the cells are bacterial cells, which, lacking eukaryotic translational and post-translational machinery, produce improperly folded TnI that must be refolded. However, any expression system can be used to produce recombinant TnI, particularly systems that require denaturation and refolding. Furthermore, this system advantageously provides an effective technique for purifying Troponin I from any source, including natural TnI and properly folded recombinant TnI under normal purification conditions (*i.e.*, under non-reducing conditions).

In accordance with the present invention there may be employed conventional molecular biology, microbiology, and recombinant DNA techniques within the skill of the art. Such techniques are explained fully in the literature. See, *e.g.*, Sambrook, Fritsch & Maniatis, *Molecular Cloning: A Laboratory Manual*, Second Edition (1989) Cold Spring Harbor Laboratory Press, Cold Spring Harbor, New York (herein "Sambrook *et al.*, 1989"); *DNA Cloning: A Practical Approach*, Volumes I and II (D.N. Glover ed. 1985); *Oligonucleotide Synthesis* (M.J. Gait ed. 1984); *Nucleic Acid Hybridization* [B.D. Hames & S.J. Higgins eds. (1985)]; *Transcription And Translation* [B.D. Hames & S.J. Higgins, eds. (1984)]; *Animal Cell Culture* [R.I. Freshney, ed. (1986)]; *Immobilized Cells And Enzymes* [IRL Press, (1986)]; B. Perbal, *A Practical Guide To Molecular Cloning* (1984); F.M. Ausubel *et al.* (eds.), *Current Protocols in Molecular Biology*, John Wiley & Sons, Inc. (1994).

The terms "express" and "expression" mean allowing or causing the information in a gene or DNA sequence to become manifest, for example producing a protein by activating the cellular functions involved in transcription and translation of a corresponding gene or DNA sequence. A DNA sequence is expressed in or by a cell to form an "expression product" such as a protein. The expression product itself, e.g. the resulting protein, may also be said to be "expressed" by the cell.

The term "expression system" means a host cell and compatible vector under suitable conditions, e.g. for the expression of a protein coded for by foreign DNA carried by the vector and introduced to the host cell. Common expression systems include *E. coli* host cells and plasmid vectors, insect host cells and *Baculovirus* vectors, and mammalian host cells and vectors. In a specific embodiment, the protein of interest is expressed in *E. coli* bacterial cells.

The term "host cell" means any cell of any organism that is selected, modified, transformed, grown, or used or manipulated in any way, for the production of a substance by the cell, for example the expression by the cell of a gene, a DNA or RNA sequence, a protein or an enzyme. Host cells can further be used for screening or other assays. Host cells can be cultured cells *in vitro* or one or more cells in a plant, e.g., a transgenic plant or a transiently transfected plant. Host cells of the invention include, though they are not limited to, bacterial cells (e.g., *E. coli*, *Synechocystis* sp., *Z. mobilis*, *Agrobacterium tumefaciens*, and *Rhodobacter*); yeast cells (e.g., *S. cerevisiae*, *Candida utilis*, *Phaffia rhodozyma*); fungi (e.g., *Phycomyces blakesleeanus*); algae (e.g., *H. pluvalis*); and plants (e.g., *Arabidopsis thaliana*).

Sulfhydryl Protecting Groups

As noted above, the sensitivity of recombinant troponin to oxidation requires maintenance of reducing conditions during purification of the protein. This results in significant drawbacks in terms of expense and difficulty.

The present invention addresses these deficiencies of the prior methods by providing sulfhydryl protected recombinant troponin I.

The term "sulfhydryl protecting group" or "cysteine protecting group" means a reversibly bound chemical group which prevents formation of intra- and intermolecular disulfide

bonds, but does not interfere with the process of protein purification. In a preferred embodiment, the “sulfhydryl protecting group” consists of sulfate groups bound through sulfitolysis with sodium tetrathionate. Numerous other reversible derivatizing reagents for cysteine sulfhydryls have been developed including disulfide compounds such as pyridyl disulfide, and the alkylalkanethiosulfonates. The sulfhydryl modification by these reagents is often facile, although their steric properties may interfere with protein activity or bioprocessing. Other sulfhydryl-reactive chemistries with potential utility in simplifying troponin I purification, recovery and storage include cyanylation and aminoethylation, reaction with compounds containing the maleimide functional group such as N-ethyl maleimide, vinyl sulfones, and alkyl halides such as iodoacetic acid and amide. However, these sulfhydryl modifying groups have the disadvantage of poor reaction reversibility, making regeneration of the free sulfhydryl sidechains of the protein cysteines more difficult.

Preferably, to ensure complete protection of Troponin I’s sulfhydryl groups and effective chromatographic purification, the Troponin I is denatured prior to reaction with the protecting groups under reducing conditions.

Chromatographic Methods

Anion exchange chromatography, hydrophobic interaction chromatography and preferably a combination thereof can be used to purify sulfhydryl protected TnI. Other suitable chromatographic techniques include cation exchange chromatography, gel permeation chromatography, reverse phase chromatography, metal chelation chromatography, etc. These chromatographic techniques can be employed in various formats, including high performance, preparative column, bulk suspension, and the like. In addition, the present invention is amenable to other standard laboratory or industrial separation techniques.

In a specific embodiment, sulfhydryl-protected recombinant TnI is purified by successive chromatographies on an ion exchange column, particularly an anion exchanger, followed by hydrophobic interaction chromatography.

Sulfhydryl-protected TnI can be stored in the protected state, either before or after chromatography. Storage of the sulfhydryl-protected TnI obviates the need for maintaining reducing

conditions, and avoids formation of intrachain or interchain disulfide crosslinks.

The sulfhydryl-protected TnI can be deprotected after chromatographic or other purification, or storage, as set forth in the following section.

TnI Deprotection

Deprotection of the sulfhydryl protected TnI can be achieved under reducing conditions, *e.g.*, to remove sulfate groups. The chemistries effective to remove reversible protecting groups are well-known in the art. Chemistry to remove these reversible sulfhydryl modifying groups often involves use of reductants such as mercaptans or dithiothreitol (Hoppe *et al.*, Biochemistry 1989, 28:2956; DiBella *et al.*, J. Biol. Chem. 1995, 270:163; Kenyon and Bruice, Methods Enzymol. 1977, 47:407; Bruice and Kenyon, J. Protein Chem. 1982, 1:47; Inoue *et al.*, Biotechnol. Appl. Biochem. 1998, 28:207).

Exemplary TnI deprotection: Desulfitolysis. Troponin may be stored or used in the sulfitolized form; if desulfitolized troponin is required the sulfhydryl group modifications may be removed by treatment of the protein with reductant such as mercaptans or dithiothreitol (DTT). At neutral to slightly acidic pH values desulfitolysis, may be accomplished by treatment of the protein with millimolar levels of reductant (see Figure 11) in the presence or absence of a solubilizing chaotrope. In normal practice, levels 10-100 fold higher are used to ensure complete conversion of the troponin. At more acidic pH values a reductant like Tris[2-carboxyethylphosphine] hydrochloride (TCEP) is preferred. The protein may be buffer exchanged by a method such as dialysis/diafiltration or gel filtration into an acidic pH buffer prior to removal of reductant to slow the formation of intermolecular disulfides.

Protein Refolding/Formulation

Purified troponin can be refolded, if desired, by first buffer exchanging the protein by dialysis, diafiltration, gel filtration or other appropriate technique into a suitable refolding buffer in the presence of a denaturing chaotrope like 8M urea or 6M guanidine. The denatured protein may then be refolded by dilution to a suitable target concentration (*e.g.*, less than 10 nanomolar) in an appropriate buffer with or without chaotrope (8-0 M urea, 6-0 M guanidine) at an appropriate

temperature, and subjected to a timed hold. Alternatively, the protein may be subjected to dialysis or diafiltration through appropriate chaotrope level transitions with hold times to promote protein refolding in a suitable refolding buffer. The refolded protein is subsequently concentrated by ultrafiltration and final formulation buffer exchange, if necessary, can be accomplished by gel
5 filtration or diafiltration.

The highly purified TnI, preferably in a refolded state, produced according to the invention can be used for any purpose, including but by no means limited to antibody generation, as a control or standard immunoassay reagent, or to inhibit angiogenesis (which can be important in treating various cancers).

EXAMPLE

The present invention will be better understood by reference to the following Example, which is provided by way of illustration and not by way of limitation.

Materials and Methods

TnI Inclusion Body Preparation. Human skeletal TnI expressed in *E. coli* was
15 isolated from lysed cells in inclusion bodies (Figure 2). To isolate and wash inclusion bodies, approximately 150 grams of cell paste was dispersed in 1.5 liter of 50 mM sodium acetate, 2 mM EDTA, pH 6. The cell suspension is subjected to two consecutive passes through a microfluidizer, 10,000 psig at 10-12°C, to break open the cells. The resultant lysate was centrifuged at 12,000 G, 4°C for 30 minutes to pellet insoluble material. The supernatant was removed and the pelleted
20 material was dispersed in 1.5 liter 50 mM sodium acetate. 2 mM EDTA, 1% Triton X-100, pH 6 and centrifuged for an additional 30 minutes at 12,000 G. The pelleted material was recovered, dispersed in 1.5 liter 50 mM sodium acetate, 2 mM EDTA, 0.5M NaCl pH 6, and centrifuged for 30 minutes at 12,000 G. The pelleted material is again dispersed in 1.5 liter of 50 mM sodium acetate, 2 mM EDTA, pH 6 and centrifuged for 30 minutes at 12,000 G. The resulting pellet is suspended
25 in 200 ml of 50 mM sodium acetate, 2 mM EDTA, pH 6 and centrifuged at 12,000 G for 30 minutes. The final pelleted inclusion bodies, approximately 10 grams, was stored at -70°C prior to subsequent processing.

Inclusion Body Solubilization and Sulfitolysis. Ten grams of TnI-containing

inclusion bodies were solubilized and protein sulfhydryls were sulfitolyzed using 200 ml 6M urea, 25 mM Tris, 10 mg/ml sodium sulfite, 5 mg/ml sodium tetrathionate pH 7.5 at ambient temperature for 6 hours in the dark (Figure 3). The solubilized material was filtered over a 0.2 micron membrane prior to subsequent processing.

5 **TnI Purification.** Sulfitolyzed recombinant human TnI was purified by a five step process (Figure 3). Solubilized, sulfitolyzed TnI-containing inclusion bodies (200 ml) were loaded onto a 3 liter volume Q-Sepharose FF (Pharmacia) column pre-equilibrated in 6M urea, 25 mM Tris, 0.1M NaCl pH 7.5 at 150 ml/minute (Figure 4). The purified TnI was collected in the column flowthrough (approximately 3000 ml, total). The recovered TnI was concentrated and buffer
10 exchanged by UF/DF using a 0.2 ft² Pall Omega cassette. Initial concentration was to 10x volume (300 ml) followed by diafiltration against 5 liters of 6M urea, 25 mM Tris pH 7.5. This material was loaded onto a 300 ml volume Q-Sepharose FF column pre-equilibrated in 6M urea, 25 mM Tris, pH 7.5 at 20 ml/minute. The bound TnI was eluted from the column by a step wash with 6M urea, 25 mM Tris, 80 mM NaCl pH 7.5 (Figures 5, 6). This eluted troponin (500 ml) was loaded onto a 60
15 ml column of Toyopearl 650M Phenyl HIC resin after addition of ammonium sulfate to a final concentration of 1M. The column was pre-equilibrated with 6M urea, 25 mM Tris, 1M ammonium sulfate pH 7.5. The purified troponin was collected as the unbound flowthrough from this column (Figures 7, 8), concentrated 2.5-fold and buffer exchanged for storage by UF/DF using a 0.2 ft² Pall Omega cartridge against 5 liters of 25 mM sodium citrate, 150 mM NaCl pH 3. Purified TnI was
20 stored frozen at -70°C.

Protein purity was determined by SDS-PAGE (Figure 8) and reverse phase chromatography (Figure 9) and protein identity was confirmed by peptide mapping with peptide mass and fragmentation analysis (Figure 10). Yield determinations for each step in the process were determined by quantitative reverse phase chromatography. Residual DNA levels, measured by DNA
25 Threshold, were less than or equal to 12 pg DNA/mg protein. Endotoxin testing of final product by LAL (gel-clot) indicated less than or equal to 3 EU/mg protein.

TnI Desulfitolyzation. For removal of sulfate groups from cysteine sidechains, purified TnI in sodium citrate storage buffer was first buffer exchanged by dialysis or diafiltration into 6M or 8M urea, 25 mM Tris, 0.15M NaCl pH 7.5. Desulfitolyzation was accomplished by

addition of DTT to a final concentration of 0.1M and incubation at ambient temperature for 45 minutes to 1 hour. Figure 11 shows the SDS-PAGE analysis of an experiment establishing the minimum conditions for desulfitolysis of recombinant human troponin I. For processing recombinant sulfitolized troponin I the level of reductant, concentration of urea and incubation time were increased over the minimum values established to insure complete conversion of the modified cysteine sidechains to the free sulfhydryl. The TnI solution was subsequently buffer exchanged by dialysis or diafiltration in the presence of reductant to a pH of 6 (8M urea, 10 mM sodium citrate, 5 mM DTT, 0.15M NaCl, pH 6), then into buffer minus reductant (8M urea, 10 mM sodium citrate, 0.15M NaCl, pH 6) to preclude the formation of intermolecular disulfides upon removal of reductant at high pH.

Analytical Methods. SDS-PAGE analysis was performed using Novex Pre-cast 16% Tris-glycine gels, sample and running buffers, and molecular weight markers. Gels were stained with Novex colloidal Coomassie blue stain.

Reverse phase chromatographic analyses were performed on an HP 1100 liquid chromatograph using a 2.1 x 150 mm Zorbax C3SB column run at a flowrate of 0.25 ml/minute at 25°C. Buffer for reverse phase column equilibration was 0.1% TFA in water. TnI samples were analyzed by gradient elution from 0.1% TFA/water to 0.1% TFA/acetonitrile at 1%/minute. Column eluates were monitored by on-line UV detection at 215 nm.

Peptide maps of TnI were generated by dilution of stock samples of TnI to 1 mg/ml with 0.1M Tris buffer, pH 8, followed by digestion with a 1:20 enzyme:substrate ratio of endoproteinase LysC at 37°C for 6 hours. LysC peptide fragments were resolved by reverse phase chromatography on a 2.1 x 150 mm Zorbax C18SB column using a gradient of 0.1% TFA/water to 0.1% TFA/acetonitrile at 0.25 ml/minute. Eluted peptide fragments of TnI were identified by on-line LC/MS detection using a Finnigan LCQ ion trap mass spectrometer set to perform automated peptide detection/fragmentation analysis.

Protein concentration analysis was performed using reverse phase analysis of TnI on the Zorbax C3SB column, whose linear detection limits were calibrated for quantitation using a TnI standard whose concentration was determined by UV 280 nm measurement and the application of the Beer/Lambert equation ($A=Ebc$) with an extinction coefficient of 0.4.

Residual DNA analysis was performed with DNA Threshold. Endotoxin contaminant was measured by Limulus Amoebocyte Lysate (LAL) gel-clot test.

Results

Samples were removed at each process step for analysis. Sample quantities were determined by reverse phase chromatography. The troponin peak absorbance at 215 nm was measured for each sample in replicate, averaged, and the quantity of protein calculated using a reverse phase calibration curve based upon a standard troponin dilution series. The calibrant standard concentration was determined using the Beer/Lambert equation, the standard absorbance at 280 nm, and an extinction coefficient of 0.4.

During final UF/DF processing, product precipitation was noted. After removing final product, residual troponin precipitate in the UF/DF cassette was resolubilized by washing with 50 ml of 6M urea, 10 mM sodium citrate, 0.15M NaCl, pH 6. This resolubilized troponin was buffer exchanged to remove urea and analyzed for troponin. The product total is the sum of the troponin recovered during the final UF/DF step and the resolubilized, buffer exchanged cassette wash.

Data Summary Table
Lot 3L5 Sulfitylized Tropinin Purification

SAMPLE	VOLUME (ML)	CONCENTRATION MG/ML	AMOUNT	RECOVERY %
Cell Pellet	-	-	148 grams	-
Inclusion Body Final Pellet	-	-	10.48 grams	-
Q1 Load, Sulfitylized Inclusion Bodies	250	2.56	640 mg	100
Q1 Flowthrough	3000	0.21	644 mg	100
Q1 UF	300	1.94	582 mg	91
Q1 DF	290	1.99	577 mg	90
Q2 Bound, 80 mM NaCl Eulate	500	0.98	489 mg	76
HIC Load, 1M Ammonium Sulfate	500	0.99	499 mg	78
HIC Flowthrough	550	0.87	479 mg	75
Final UF/DF	200	1.60	318 mg	50
Cassette Wash/Reprocessed Precipitate	50	0.69	34 mg	5
Product Total	-	-	352 mg	55

* * *

The present invention is not to be limited in scope by the specific embodiments described herein. Indeed, various modifications of the invention in addition to those described herein will become apparent to those skilled in the art from the foregoing description and the accompanying figures. Such modifications are intended to fall within the scope of the appended claims.

It is further to be understood that all values are approximate, and are provided for description.

All patents, patent applications, publications, procedures, and other materials cited herein are hereby incorporated by reference in their entireties.